



Distribution of bacterioplankton lineages within the dark ocean of the Sargasso Sea and their potential role in biogeochemical cycles

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Overview

Marine bacterioplankton (bacteria and archaea) are recognized for their importance in global biogeochemical transformations. Studying their spatial and temporal distribution is necessary to understanding oceanic nutrient cycling.

Table 1. Selected bacterial and archaeal bacterioplankton lineages for this study and their metabolic potential.

SAR11	Photoheterotrophy: proteorhodopsin ^{1,2}
SAR202	Oxidation of recalcitrant organic compounds (FMNOs) ³
SAR324	Chemoautotrophy: carbon fixation (cbbL) ⁴
Euryarchaeota	Methanogenesis (mcrA) ⁵ , proteorhodopsin ⁶
Thaumarchaeota	Chemoautotrophy: ammonia oxidation (amoA) ⁷

The Bermuda Atlantic Time-series site (BATS) offers an opportunity to study bacterioplankton communities with biogeochemical variables in the context of the oxygen minimum.

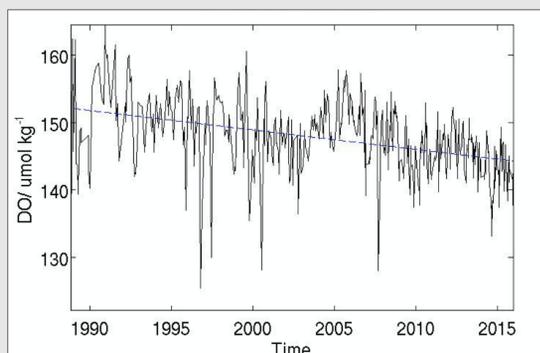


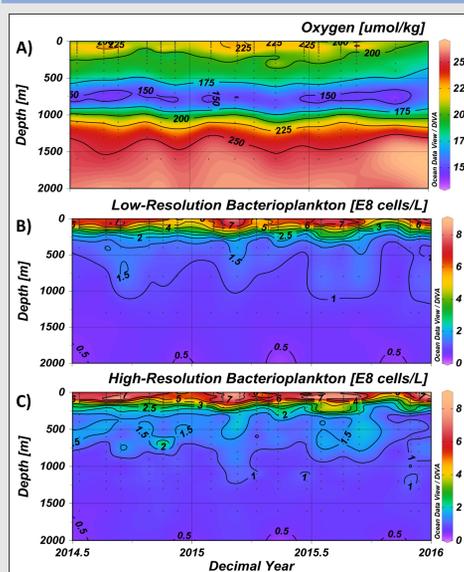
Fig. 1. Time series of the oxygen minimum in $\mu\text{mol}/\text{kg}$ at BATS from 1989 to 2016.

With cellular respiration by heterotrophic bacteria and archaea fueling the oxygen minimum (Fig. 2), studying the metabolic processes within this zone will contribute to our understanding of the ongoing decrease in dissolved oxygen concentrations (Fig. 1) and vertical expansion of the oxygen minimum zone at BATS.

Methods

<p>1</p> <p>Sampling</p> <p>CTD Rosette aboard the R/V <i>Atlantic Explorer</i></p> <p>BATS site 31°40' N 64°10' W 0-4500 m depth</p> <p>July 2014 to December 2015</p>	<p>2</p> <p>Epifluorescent microscopy</p> <p>DAPI stain: total bacterioplankton</p> <p>FISH*: SAR202</p> <p>CARD-FISH**: SAR11, SAR324 & archaea</p> <p>40 mL samples fixed with 10% formalin</p> <p>* Fluorescence <i>in situ</i> hybridization ** Catalyzed reporter deposition fluorescence <i>in situ</i> hybridization</p>	<p>3</p> <p>Gene analysis</p> <p>Phenol-chloroform DNA isolation method</p> <p>amoA & cbbL amplified by optimized PCR method</p>
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Bacterioplankton contribute to the oxygen minimum



The distribution of total bacterioplankton was examined at high vertical resolution in the context of the low oxygen realm (Fig. 2A) revealing a subsurface maximum that negatively correlates with oxygen levels (Table 2).

Table 2. Correlation between oxygen levels and total bacterioplankton counts.

Depth	0-400 m	400-2000 m
r	0.6823	-0.6848
p	<0.001	<0.001
n	153	122

Fig. 2. A) Oxygen levels, B) bacterioplankton abundance at low-resolution sampling and C) bacterioplankton abundance at high-resolution sampling from Jul. 2014 to Dec. 2015.

Spatial and temporal distribution of select lineages

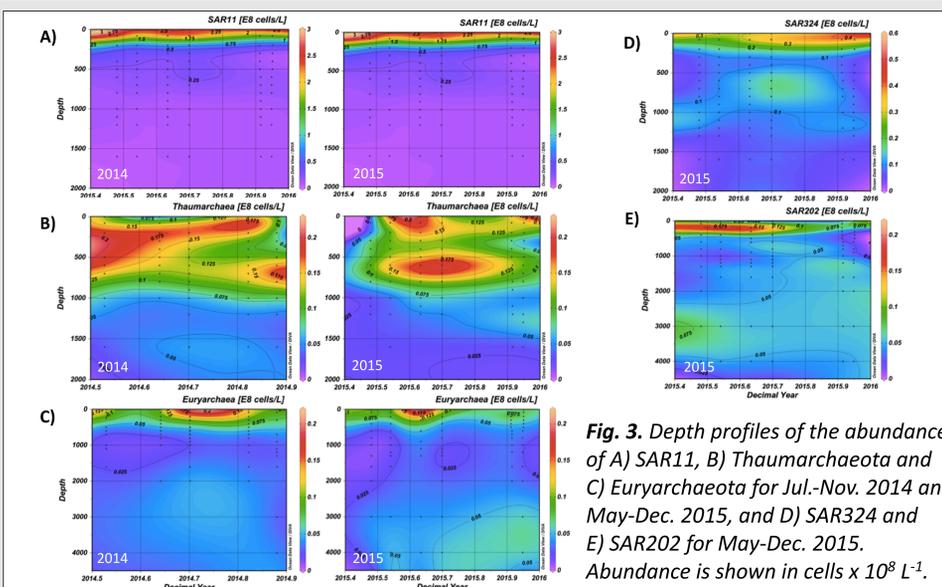


Fig. 3. Depth profiles of the abundance of A) SAR11, B) Thaumarchaeota and C) Euryarchaeota for Jul.-Nov. 2014 and May-Dec. 2015, and D) SAR324 and E) SAR202 for May-Dec. 2015. Abundance is shown in $\text{cells} \times 10^8 \text{ L}^{-1}$.

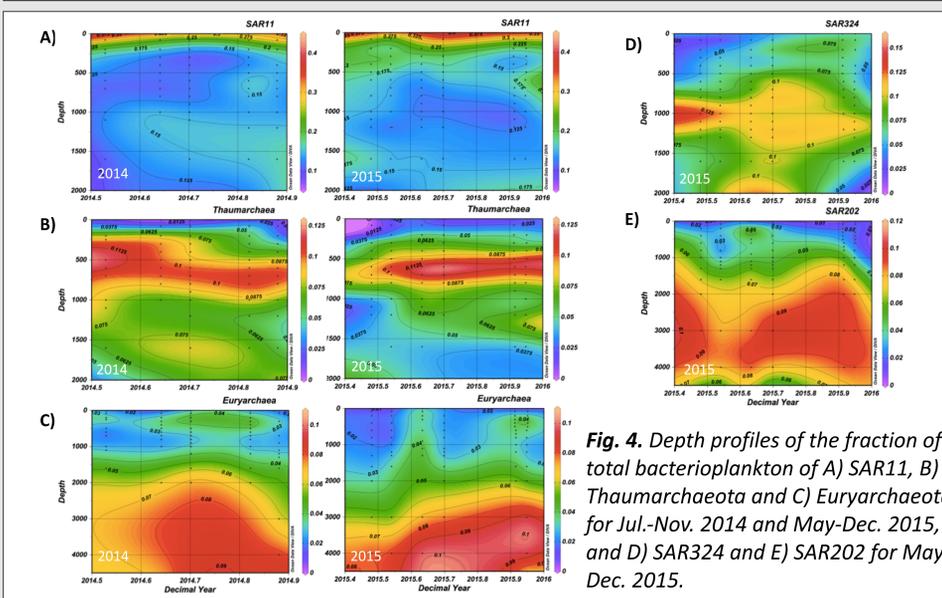


Fig. 4. Depth profiles of the fraction of total bacterioplankton of A) SAR11, B) Thaumarchaeota and C) Euryarchaeota for Jul.-Nov. 2014 and May-Dec. 2015, and D) SAR324 and E) SAR202 for May-Dec. 2015.

Nitrification and carbon fixation

Genes for archaeal ammonia oxidation (*amoA*: ammonia monoxygenase) and carbon fixation (*cbbL*: form I RubisCO) associated with *Thaumarchaeota* and SAR324, respectively, were investigated throughout the water column^{7,8}.

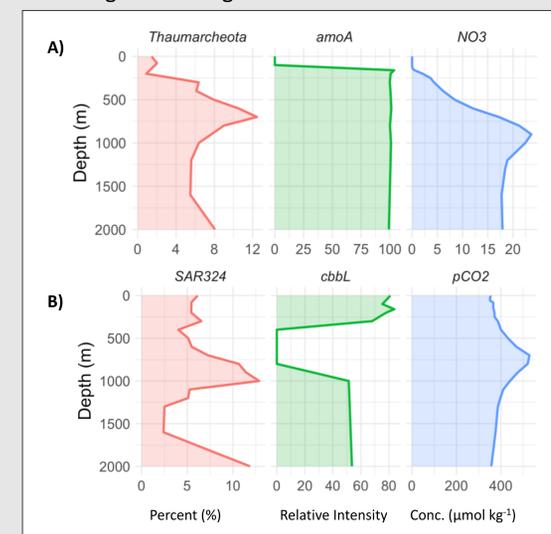


Fig. 5. A) *Thaumarchaeota* percent contribution, *amoA* gene intensity and nitrate concentrations and B) SAR324 percent contribution, *cbbL* gene intensity and $p\text{CO}_2$ levels for Nov. 2014.

Conclusion

Bacterioplankton respiration contributes to the oxygen minimum.

SAR11 abundance and percent contribution are highest in surface and mesophotic waters ($\sim 0-400$ m).

Thaumarchaeota abundance and percent contribution are highest in the upper oxycline ($\sim 400-800$ m) and show a relationship with **archaeal ammonia oxidation**.

SAR324 contributes the most to the lower oxycline ($\sim 800-1200$ m) bacterioplankton community and show correlation with **carbon fixation**.

Euryarchaeota & SAR202 contribute the most to the deep ocean (>2000 m) bacterioplankton community.

Future Directions

- Further *amoA* and *cbbL* depth profile analysis through qPCR.
- Illumina sequencing of 16S rRNA gene amplicons.
- Lineage abundance and diversity data will be put into the context of the biogeochemical data provided at BATS.

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